Interaction of aluminum ion with ATP. Mechanism of the aluminum inhibition of glycerol kinase and its reversal by spermine

Masataka Yoshino, Keiko Murakami & Keiichi Kawano*

Department of Biochemistry, Aichi Medical University, Nagakute, and *Division of Biological Sciences, Graduate School of Science, Hokkaido University, Sapporo, Japan.

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Aluminum ion inhibited yeast glycerol kinase competitively with respect to the substrate MgATP. The K_i value of the enzyme for aluminum ion was about 3 µM. Spermine at physiological concentrations prevented glycerol kinase from the inhibition by aluminum ion. Nuclear magnetic resonance spectroscopy showed the specific elimination by spermine of aluminum from the metal-ATP complex, but no dissociation of MgATP complex by spermine. Inhibition by aluminum ion of glycerol kinase as well as hexokinase can reduce the utilization of energy fuel in yeast. Change in polyamine concentration may control energy production in vivo, and is responsible for the development of age-related aluminum toxicity.

Keywords: aluminum ion, glycerol kinase, NMR, spermine, yeast

Introduction

Aluminum is the most abundant metal in the earth's crust, but is present in small amounts in living organisms because of its low solubility at neutral pH (Martin 1986, Macdonald & Martin 1988). Recent reports show aluminum toxicity on various biochemical reactions (Martin 1986, Macdonald & Martin 1988, Ganrot 1986). Ionic radius for Al3+ most closely resembles those of Mg^{2+} and Fe^{3+} , and thus, Al3+ substitution for Mg2+ at critical target sites causes the inhibition of several Mg²⁺-dependent enzymes by aluminum (Martin 1986, Macdonald & Martin 1988). Potent inhibition by aluminum ion of hexokinase (EC 2.7.1.1) (Lai & Blass 1984, Womack & Colowick 1979, Yoshino et al. 1990) and NADPlinked isocitrate dehydrogenase (EC 1.1.1.42) (Yoshino et al. 1990, Yoshino & Murakami 1992, Yoshino et al. 1992) may contribute to the aluminum toxicity in cells.

Address for correspondence: M. Yoshino, Department of Biochemistry, Aichi Medical University, Nagakute, Aichi 480-11, Japan. Fax: (0561) 61 4056; E-mail: yoshino@aichi-med-u.ac.jp

In this paper we report the inhibitory effect of aluminum ion on the activity of glycerol kinase (EC 2.7.1.30), the first enzyme of the glycerol-utilizing pathway in yeast. The inhibition of glycerol kinase by aluminum was effectively reversed by spermine the principal polyamine in eukaryotes. Nuclear magnetic resonance (NMR) studies revealed that spermine specifically ejected aluminum ion from the metal-ATP complex, but that spermine did not affect the binding of Mg2+ to ATP. Inhibition of glycerol kinase by aluminum ion and its reversal by spermine may be related to the appearance of aluminum toxicity in cells.

Materials and methods

NADP, purified glycerol kinase (specific activity of 160 µmol min-1 per mg protein) from yeast (Candida mycoderma) and rabbit muscle glycerol 3-phosphate dehydrogenase (EC 1.1.1.8, specific activity of 320 µmol min⁻¹ per mg protein) were obtained from Boehringer-Mannheim-Yamanouchi (Tokyo, Japan). 4-Morpholinopropanesulfonic acid (Mops) was obtained from Dojindo Co. (Kumamoto, Japan).

Kinetic studies

Glycerol kinase activity was determined by following the change in absorbance at 340 nm. The assay medium of 0.5 ml contained 50 mM Mops–KOH buffer (pH 6.85), various concentrations of $MgCl_2$ and ATP, 4 mM glycerol and the enzyme in the absence and presence of $AlCl_3$. The reaction was started by adding the enzyme. After incubation at 37 °C for 5 min, the reaction was stopped by addition of 60 μ l of 100 mM EDTA and 0.98 ml of 0.2 m glycine–KOH buffer (pH 9.8) containing 200 mg ml⁻¹ hydrazine hydrate. NAD and purified glycerol 3-phosphate dehydrogenase were added to final concentrations of 0.1 mM and 10 units ml⁻¹, respectively. The mixture was incubated at 37 °C for 10 min, and the absorbance at 340 nm was determined.

Measurement of ²⁷Al- and ²⁵Mg-NMR spectra

 $^{27}\mbox{Al-NMR}$ and $^{25}\mbox{Mg-NMR}$ spectra were recorded at 104.26 MHz and 24.48 MHz, respectively, with a Bruker AM400 spectrometer. An equipped transmitter provided 90-degree pulse width of 46 μs for $^{25}\mbox{Mg}$ and 17 μs for $^{27}\mbox{Al}$, respectively. Typical $^{27}\mbox{Al}$ spectra consists of collecting 320 to 960 transients using 16K data points over 10 000 Hz spectral width. Measurements of $^{25}\mbox{Mg}$ spectra were performed with the same condition of $^{27}\mbox{Al}$ using 960 to 56 000 transients. The signal to noise ratio was improved by exponential multiplication which introduced 1 to 10 Hz line broadening.

Results

Addition of Al^{3+} inhibited the yeast glycerol kinase with a decrease in the affinity for the substrate MgATP (Figure 1(A)). Double reciprocal plot revealed the inhibition to be of a competitive type with respect to MgATP (Figure 1(B)). Figure 2 shows the effect of increasing concentrations of Al^{3+} on the glycerol kinase activity. Dixon plot showed that Al^{3+} acted as a competitive inhibitor of the enzyme with a K_i value of 3 μ M (Figure 2(B)).

Spermine effectively protects hexokinase from the inhibition by aluminum ion (Yoshino *et al.* 1990). Thus, we examined the effect of spermine on the aluminum-induced inhibition of glycerol kinase. Apparent K_i values of the enzyme for aluminum ion increased with the increase in spermine concentration: addition of spermine at physiological concentration reduced the affinity of the enzyme for aluminum to half (Figure 3).

Reversal of aluminum-inhibition by spermine suggests that spermine inhibits the formation of the aluminum-ATP complex. We analyzed the action of spermine on the complex formation between aluminum and ATP using NMR spectroscopy.

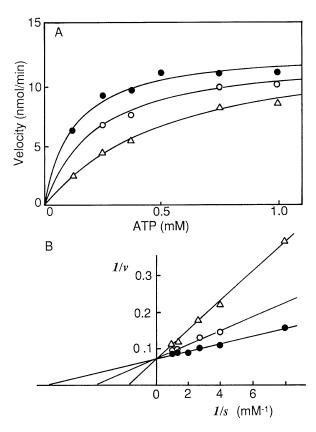


Figure 1. Effect of the concentrations of threo-Ds-isocitrate on the activity of yeast glycerol kinase in the absence and presence of aluminum ion. The reaction mixture of 0.5 ml contained 50 mm Mops-KOH buffer (pH 6.85), various concentrations of MgCl₂ and ATP, 4 mm glycerol and the enzyme in the absence and presence of AlCl₃. MgCl₂ was added to obtain the indicated concentrations of MgATP. The reaction was started by adding the enzyme. After incubation at 37 °C for 5 min, glycerol 3phosphate formed was determined enzymatically as described in Materials and methods. (A) MgATP saturation curves in the absence (\bullet) and presence of 5 μ M (\bigcirc) or 20 μ M (\triangle) aluminum ion. (**B**) Double reciprocal plot. Points represent experimental data, and lines are theoretically drawn according to the non-linear regression analysis (Duggleby 1981).

Figure 4(A) shows the ²⁷Al-NMR spectra with a sharp singlet resonance of aluminum ion at pH 2. Addition of ATP at twice the concentration of aluminum ion decreased the sharp resonance of Al³⁺ markedly, and a broad signal appeared at 4 p.p.m. higher magnetic field than free aluminum ion (Figure 4(B)). We assigned this resonance to the aluminum–ATP complex (Karlik *et al.* 1982). The broadening of the resonance results from a decrease in the symmetry of the ligand field as a result of the substitution of P for H₂O in the Al³⁺ coordination

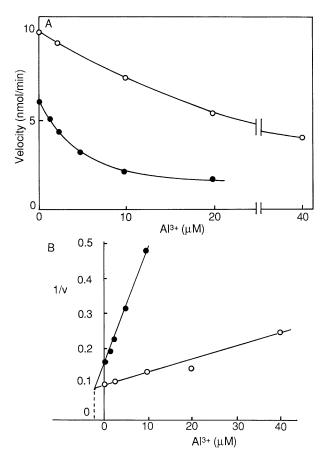


Figure 2. Effect of aluminum ion on the activity of yeast glycerol kinase in the presence of different concentrations of MgATP. Reaction mixture was similar to that in legend to Figure 1, except that AlCl₃ concentrations were varied. (A) Inhibition curves. (B) Dixon plot. (●) 0.125 mM MgATP. (O) 0.5 mm MgATP.

sphere. Further addition of excess spermine to the aluminum-ATP complex increased the intensity of the sharp resonance of free aluminum ion (Figure 4(C)). Increase in spermine concentration caused a further increase in the free aluminum ion signal with concomitant reduction of broad resonance of ATPaluminum complex (Figure 4(D)). These results indicate that spermine can eject Al3+ from the aluminum-ATP complex.

We further explored the effect of spermine on the MgATP complex using ²⁵Mg-NMR spectroscopy. Magnesium ion gave a sharp singlet resonance (Figure 5(A)), but addition of ATP caused a marked broadening and reduction of the signal intensity (Figure 5(B)). ²⁵Mg nuclei has a large quadrupolar moment and the broad resonance is also due to the coordinated H₂O (Shimizu & Hatano 1982). Further addition of spermine to the MgATP complex could

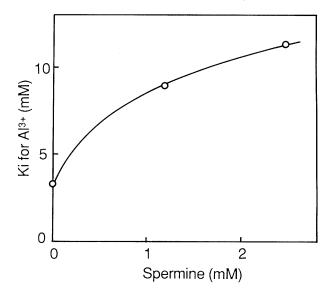


Figure 3. Effect of spermine on the K_i values of glycerol kinase for aluminum ion. Reaction mixture was similar to that in Figure 2, but spermine was included as indicated.

not recover the sharp resonance of Mg²⁺ (Figure 5(C)). Thus, spermine can specifically eject aluminum ion from the metal-ATP complex, but not magnesium ion from the complex.

Discussion

Aluminum, the most abundant metal in the environment, shows various toxic effects on biological and biochemical processes, but the molecular mechanism of aluminum toxicity has remained obscure. The toxic effect of aluminum may be the substitution for some essential metal ions at critical sites in the cells (Ganrot 1986). Binding of metal ion is primarily electrostatic and therefore, in addition to charge, ionic size is an important parameter. Ionic radius for Al³⁺ most closely resembles those of Mg²⁺ and Fe³⁺, and Al³⁺ would be substituted for these ions under physiological conditions (Martin 1986, Macdonald & Martin 1988). Thus, aluminum ion can act as an inhibitor of several Mg²⁺-dependent enzymes in cells (Ganrot 1986). Typical examples are the inhibition of hexokinase (Womack & Colowick 1979, Lai & Blass 1984, Yoshino et al. 1990) and NADP-isocitrate dehydrogenase (Yoshino et al. 1992, Yoshino & Murakami 1992), which results in the inhibition of the energy-yielding reaction in cells. Aluminum inhibition of these enzymes may be responsible for some neurological and skeletal disorders (Ganrot 1986). Furthermore, inhibitory action of aluminum ion on

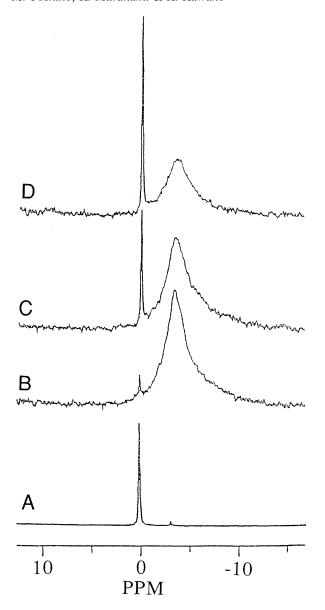


Figure 4. ²⁷Al–NMR spectra of aluminum ion and aluminum–ATP complex in the absence and presence of spermine. pH of the solution was kept at 2.0. (**A**) 1 mM AlCl₃. (**B**) 2 mM ATP plus 1 mM AlCl₃. (**C**) 20 mM spermine, 2 mM AlCl₃ and 1 mM ATP. (**D**) 140 mM spermine, 2 mM AlCl₃ and 1 mM ATP.

these enzymes may participate in the inhibition of root elongation of plants causing forest decline (Godbold *et al.* 1988, Yoshino *et al.* 1992).

The present results show that aluminum ion inhibits glycerol kinase in a competitive manner with respect to the substrate MgATP. Effect of aluminum ion on the enzyme was similar to the inhibition of hexokinase (Womack & Colowick 1979, Lai & Blass 1984). Aluminum ion forms a chelate complex of

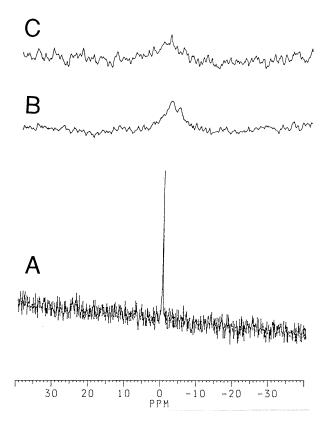


Figure 5. ²⁵Mg–NMR spectra of magnesium ion and MgATP complex in the absence and presence of spermine. (**A**) 10 mM MgCl₂ at pH 6.25. (**B**) 10 mM MgCl₂ plus 10 mM ATP at pH 7.20. (**C**) 10 mM MgCl₂ plus 10 mM ATP in the presence of 50 mM spermine.

aluminum–ATP, and can bind to the MgATP-sites of these enzymes. The $K_{\rm i}$ values of these two enzymes for aluminum ion were similar to each other: 3 and 1.8 μ M for glycerol kinase and hexokinase, respectively. Furthermore, spermine, which is not a direct activator of these enzymes, reversed the inhibition by aluminum ion. NMR spectroscopy showed that spermine specifically eliminates aluminum ion from the aluminum–ATP complex, but cannot eject Mg²⁺ from the MgATP complex can explain the reversal by spermine of aluminum inhibition of Mg²⁺-dependent enzymes.

Glycerol kinase and hexokinase utilize glycerol and glucose, respectively, as an energy source by phosphorylation and subsequent glycolysis or oxidation in yeast (Thorner & Paulus 1973). The inhibition by aluminum ion of these enzymes can suppress the energy production in yeast. Spermine concentration necessary for the reversal of the aluminum inhibition of glycerol kinase and hexokinase (Yoshino *et al.* 1990) is about 1 to 2 mM, within

physiological range (Abrahams & Pihl 1981). Spermine the principal polyamine responsible for the macromolecular synthesis accumulates with cell proliferation in cells, whereas aging of cell decreases polyamines (Abrahams & Pihl 1981). Decrease in polyamine can enhance the inhibition by aluminum ion of these enzymes, resulting in the reduction of the phosphorylation of glycerol and glucose. Aluminum toxicity may be explained by the inhibition of these enzymes, and further by the decreased reversal of the inhibition resulting from the decreased polyamine.

References

- Abrahams AK, Pihl A. 1981 Role of polyamines in macromolecular synthesis. Trends Biochem Sci 6, 106-107.
- Duggleby RG. 1981 A nonlinear regression analysis for small computers. Anal Biochem 110, 9–18.
- Ganrot PO. 1986 Metabolism and possible health effects of aluminum. Environ Health Perspect 85, 363-441.
- Godbold DL, Fritz E, Huttermann A. 1988 Aluminum toxicity and forest decline. Proc Natl Acad Sci USA 85, 3888-3892.

- Karlik SJ, Elgavish GA, Pillai RP, Elichhorn GL. 1982 Aluminum-27 NMR studies of Al(III)-phosphate complexes in aqueous solution. J Magn Reson 49, 164–167.
- Lai JCK, Blass JP. 1984 Inhibition of brain glycolysis by aluminum. J Neurochem **42**, 438–446.
- Macdonald T, Martin RB. 1988 Aluminum ion in biological system. Trends Biochem Sci 13, 15–19.
- Martin RB. 1986 The chemistry of aluminum as related to biology and medicine. Clin Chem 32, 1797–1806.
- Shimizu T, Hatano M. 1982 ²⁵Mg-NMR study of Mg²⁺-ATP, ADP-creatine kinase complexes. Biochem Biophys Res Commun 104, 720-726.
- Thorner JW, Paulus H. 1973 Glycerol and glycerate kinases. In: Boyer DP, ed. The Enzymes, 3rd edn. New York: Academic Press; 487-508.
- Womack FC, Colowick SP. 1979 Proton-dependent inhibition of yeast and brain hexokinases by aluminum in ATP preparations. Proc Natl Acad Sci USA 76, 5080-5084.
- Yoshino M, Murakami K. 1992 Aluminum: a pH-dependent inhibitor of NADP-isocitrate dehydrogenase from porcine heart. BioMetals 5, 217-221.
- Yoshino M, Murakami K, Yamada Y. 1990 Reversal by polyamine of the aluminum-induced inhibition of hexokinase from human brain. Biomedical Res 11, 215–218.
- Yoshino M, Yamada Y, Murakami K. 1992 Inhibition by aluminum ion of NAD- and NADP-dependent isocitrate dehydrogenases from yeast. Int J Biochem 24, 1615-1618.